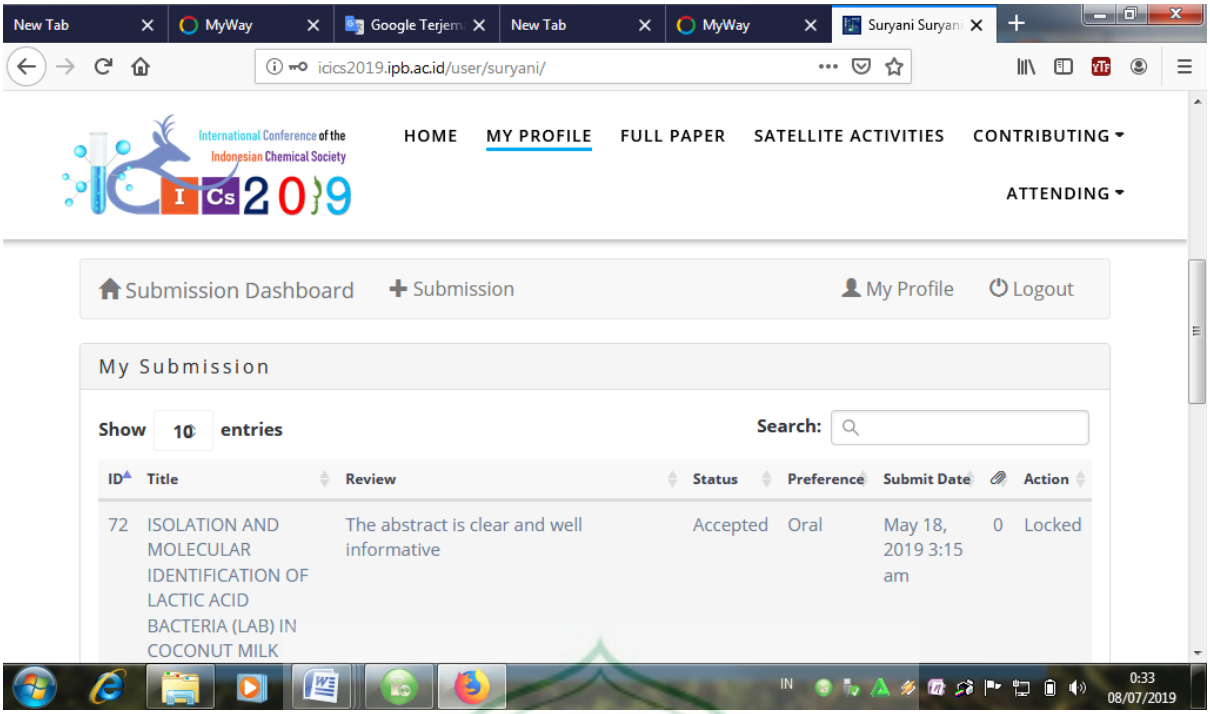

Symposium Program

Day 1: Tuesday, August 6th, 2019
Venue: IPB International Convention Center (IICC), Botani Square, Jl. Pajajaran, Bogor, West Java Indonesia

Time	Agenda		
08.00–08.45	Registration		
08.45–09.10	Welcoming Speech		
09.10–09.15	Opening Ceremony		
09.15–09.30	*Coffee Break*		
09.30–09.55	**Plenary session** Invited Speaker session: Prof Tsuyoshi Kawai		
09.55–10.20	**Plenary session** Invited Speaker session: Assoc Prof Yuan-Chung Cheng		
10.20–10.35	Discussion		
10.35–11.00	**Plenary session** Invited Speaker session: Prof Masaki Kita		
11.00–11.25	**Plenary session** Invited Speaker session: Prof Dr Dyah Iswantini		
11.25–11.40	Discussion		
11.40–12.10	Technical Presentation		
12.10–13.15	*Lunch*		
13.15–14.00	Poster Session I and Sponsor Stand Tour		
14.00–15.30	**Parallel session**		
	Ballroom 1 Invited Speaker: Shuichi Shinma, Ph.D OP A1-A5	Ballroom 2 Invited Speaker: Akhmad Sabarudin, D.Sc OP B1-B5	Meeting Room B Invited Speaker: Prof Lee Wah Lim OP C1-C5
	Meeting Room C Invited Speaker: Novriyandi Hanif, D.Sc OP D1-D5	Meeting Room D Invited Speaker: Prof Asep Kadarohman OP E1-E5	Meeting Room E Invited Speaker: Dr. Dodi Safari OP F1-F5

GUIDELINES

All participants

1. Please attend the entire conference program, so that speaker will not be disappointed because of small audience. Conference is to hear and exchange ideas.
2. Please do not smoke and talk in the room
3. Please silent your mobile phone
4. Please do not take photograph and/or video during the presentation
5. Participants are welcome to choose parallel rooms and please ask only short and clear questions
6. Certificates will be given at the end of the symposium

Oral Presenter

1. Presentation time is scheduled by the committee
2. The room will be equipped with a LCD projector, and an official PC. The committee does not recommend the presenter to use a personal computer. Presentation file should be written in Microsoft Power Point
3. The committee prepare the rehearsal time for you before your presentation time (please see the rehearsal schedule). You can check either your presentation file is fine or need to revise to fit with the equipment.
4. Rehearsal time for oral presenter

Oral presenter A1 – A10	6 August 2019, 13.00 – 13.15 at ballroom 1
Oral presenter A6 – A10	6 August 2019, 15.40 – 15.50 at ballroom 1
Oral presenter B1 – B10	6 August 2019, 13.00 – 13.15 at ballroom 2
Oral presenter B6 – B10	6 August 2019, 15.40 – 15.50 at ballroom 2
Oral presenter C1 – C10	6 August 2019, 13.00 – 13.15 at B room
Oral presenter C6 – C10	6 August 2019, 15.40 – 15.50 at B room

ORAL PRESENTATION SCHEDULE

Name	ID submision	Code	Day	Room	Time
Adam Wiryawan	358	H2	6 Agt	Adenium Room	14.10-14.20
Agung Bagus Pambudi	328	B16	7 Agt	Ballroom 2	13.30-13.40
Agus Abhi Purwoko	29	G9	6 Agt	Room F	16.10-16.20
Agus Dwi Ananto	50	H17	7 Agt	Adenium Room	11.10-11.20
Aisy Rifa Cahyani	233	I14	7 Agt	Plumeria Room	10.30-10.40
Alex L. Suherman	389	B21	7 Agt	Ballroom 2	14.30-14.40
Alfiyatul Fithri	131	I9	6 Agt	Plumeria Room	16.10-16.20
Aliya Nur Hasanah	28	A1	6 Agt	Ballroom 1	14.30-14.40
Alvin Rahmad Widyanto	276	C11	7 Agt	Room B	10.50-11.00
Amanda Dwikarina	264	D17	7 Agt	Room C	13.40-13.50
Ani Iryani	347	B18	7 Agt	Ballroom 2	13.50-14.00
Annisa Indriyani	363	F21	7 Agt	Room E	14.30-14.40
Antonius Padua Ratu	42	H14	7 Agt	Adenium Room	10.30-10.40
Antuni Wiyarsi	366	H7	6 Agt	Adenium Room	15.10-15.20
Anugrah Ricky Wijaya	349	D20	7 Agt	Room C	14.30-14.40
Ari Asnani	351	F20	7 Agt	Room E	14.20-14.30
Arif Rahman	329	C18	7 Agt	Room B	13.50-14.00
Asdim	378	C19	7 Agt	Room B	14.00-14.10
Asri S. Mahulette	32	H9	6 Agt	Adenium Room	16.10-16.20
Atika Oktrima Puspa	254	F12	7 Agt	Room E	11.00-11.10
Awan Rahmadewi	277	F16	7 Agt	Room E	13.30-13.40
Baiq Desy Ratnasari	62	D19	7 Agt	Room C	14.20-14.30
Bambang Piluharto	310	I19	7 Agt	Plumeria Room	11.30-11.40
Bambang Purwono	344	E10	6 Agt	Room D	16.40-16.50
Betty Marita Soebrata	332	C22	7 Agt	Room B	14.40-14.50
Budi Arifin	71	E9	6 Agt	Room D	16.30-16.40
Budi Riza Putra	9	G14	7 Agt	Room F	10.30-10.40

Name	ID submision	Code	Day	Room	Time
Siti Nurbayti	268	G20	7 Agt	Room F	13.30-13.40
Sri Kadarwati	346	I17	7 Agt	Plumeria Room	11.10-11.20
Sri Mulijani	161	I10	6 Agt	Plumeria Room	16.20-16.30
Sri Mulijani	337	I11	6 Agt	Plumeria Room	16.30-16.40
Sri Sugiarti	73	G11	6 Agt	F Room	16.30-16.40
Sri Yadial Chalid	8	F1	6 Agt	Room E	14.30-14.40
Sry Wahyuni	67	B3	6 Agt	Ballroom 2	14.50-15.00
Sry Wahyuni	205	B8	6 Agt	Ballroom 2	16.20-16.30
Subandi	375	F22	7 Agt	Room E	14.40-14.50
Sudirman	43	I8	6 Agt	Plumeria Room	16.00-16.10
Suharso	15	C1	6 Agt	Room B	14.30-14.40
Suryani	72	F5	6 Agt	Room E	15.10-15.20
Sutrisno	368	B22	7 Agt	Ballroom 2	14.40-14.50
Syafrizayanti	27	F2	6 Agt	Room E	14.40-14.50
Tanto Budi Susilo	118	H15	7 Agt	Adenium Room	10.50-11.00
Tatas H.P. Brotosudarmo	352	D23	6 Agt	Plumeria Room	16.40-16.50
Teguh Pambudi	122	C5	6 Agt	Room B	15.10-15.20
Triana Kusumaningsih	33	E8	6 Agt	Room D	16.20-16.30
Trianda Ayuning Tyas	105	G13	7 Agt	Room F	10.20-10.30
Uswatun Hasanah	153	B1	6 Agt	Ballroom 2	14.30-14.40
Verra Nurmaylindha	134	H22	7 Agt	Adenium Room	13.50-14.00
Vina Juliana	309	D21	7 Agt	Room C	14.40-14.50
Wahyu Prasetyo Utomo	299	E22	7 Agt	Room D	14.40-14.50
Waringin Margi Yusmaman	144	G24	7 Agt	Room F	14.30-14.40
Wega Trisunaryanti	74	H10	6 Agt	Adenium Room	16.20-16.30
Widia Wati	133	H21	7 Agt	Adenium Room	13.40-13.50
Wilis Okti Pamungkas	338	F19	7 Agt	Room E	14.00-14.10
Winda Andika	355	E6	6 Agt	Room D	16.00-16.10

ISOLATION AND MOLECULAR IDENTIFICATION OF LACTIC ACID BACTERIA (LAB) IN COCONUT MILK FERMENTATION

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ABSTRACT

Lactic Acid Bacteria (LAB) contains of bacteriocin which is peptide that has the capacity to isolate the growth of pathogenic bacteria, where in contrast is harmless for other good bacteria. LAB is found in material fermentation containing high carbohydrate and protein like coconut milk which is undergone the process becoming Virgin Coconut Oil (VCO). In the fermentation process, there were three layers formed; oil, blondo, and water (waste). The LAB isolation on coconut milk fermentation used MRSA + 0,5% $CaCO_3$ as the selective media. with the dilution from 10^{-1} to 10^{-7}. Here, each sample was taken from each layer formed in the milk fermentation process. The identification was carried out in two ways, first was morphology identification, and the second one was molecular identification applying the PCR method. There were 97 isolates obtained from oil layer, 23 isolates from Blondo layer, and 14 isolates from water layer. After being identified well based on both morphology, and molecular on the oil layer, there were six LAB found, which were *Lactobacillus paracasei, Lactobacillus plantarum, Micrococcus luteus, Corineaebacterium bovis, Lactobacillus thermobacterium* dan *Corineaebacterium xerocis.* Three types of LAB within the blondo were identified as *Lactobacillus plantarum, Lactobacillus paracasei* and *Lactobacillus thermobacterium.*

Keywords: Isolation, molecular identification, Virgin Coconut Oil (VC)_, Lactic Acid Bacteria (LAB), PCR

Day 1: Tuesday, August 6th, 2019
Room Paralel 1 (Ballroom 1)

Time	ID/ Code	Presenter & Title	
14.30-14.40	28/A1	Aliya Nur Hasanah	DIAZEPAM MOLECULAR IMPRINTED POLYMER SOLID PHASE EXTRACTION (MI-SPE) WITH ACRYLAMIDE AND METHYL METHACRYLATE AS FUNCTIONAL MONOMER
14.40-14.50	58/A2	Lelifajri	Study on Methylene Blue Dye Adsorption in Aqueous Solution by Heat-treated Gnetum gnemon shell waste particles as Low-Cost Adsorbent
14.50-15.00	61/A3	RIFKI HUSNUL KHULUK	SIMULTANEOUS DETERMINATION OF SOME FLAVONOIDS AND CLASSIFICATION OF DIFFERENT PLANT PARTS AND GEOGRAPHICAL ORIGIN OF Sonchus arvensis
15.00-15.10	86/A4	Donatus Rendo	Removal of Methylene Blue Dye in Water by Using Separable Natural Zeolite/Fe3O4 Adsorbent
15.10-15.20	106/A5	Hasmalina Nasution	The Effect Of Using Durian (Durio zibethinus Murr) Seed Flour On Patin Fish (Pangasius hypophthalmus) Nugget Nutrient
16.00-16.10	108/A6	Muh. Supwatul Hakim	Optical chemical sensor based on incorporation of 2,2 furil dioxime in sol-gel matrix for determination of Ni (II) in water
16.10-16.20	138/A7	Muhammad Bakhru Thohir	OPTICAL SENSOR FOR NICKEL BASED ON THIN FILMS OF SOL-GEL/PAPER WITH TEOS PRECURSOR AND LIGAN İ±-FURILDIOXIME
16.20-16.30	168/A8	Erin Ryantin Gunawan	Separation of The Fatty Acid Ethanolamides Component Using High Performance Liquid Chromatography
16.30-16.40	169/A9	Dedy Suhendra	Lipase Catalyzed Production of N-Methyl Fatty Hydroxamic Acids from Terminalia catappa L. Seed Oil
16.40-16.50	171/A10	Refilda	Determination of Antioxidant in Fermented Red Betel Leaf Extract (Piper crocatum) and Its Effect on Red Chili Growth (Capsicum annuum L.)

15.20		Sinambela	Vanadyl β-diketonate Complexes
16.00-16.10	355/E6	Winda Andika	Dammarane-type Triterpenoid from The Stem Bark of Aglaia elaeagnoidea (A.juss) Benth (Meliaceae)
16.10-16.20	24/E7	Jufrizal Syahri	QSAR STUDY ON FLUOROQUINOLONE DERIVATIVES AS POTENTIAL ANTIBACTERIAL AGENTS
16.20-16.30	33/E8	Triana Kusumaningsih	An efficient and greener synthesis of 2, 4-diacetylphloroglucinol catalyzed by sulphuric acid adsorbed on silica gel and its environmental assessment
16.30-16.40	71/E9	Budi Arifin	Synthesis of C-Prenylated 1,3-Diketone Intermediate of 3-Prenylflavone
16.40-16.50	344/E10	Bambang Purwono	SYNTHESIS AND ACTIVITY ASSAY OF BENZIMIDAZOLE DERIVATIVES AS AN ANTIMALARIAL

Room Paralel 6 (Meeting Room E)

Time	ID/ Code	Presenter & Title	
14.30-14.40	8/F1	Sri Yadial Chalid	Profil of Peanut (Arachis hypogaea L.) Protein Extract as the Reagents of Allergy Test with Skin Prick Test (SPT) Method
14.40-14.50	27/F2	Syafrizayanti	In vitro cytotoxity of 3-Oxoolean-12-en-27-oic acid compound isolated from Sandoricum koetjape Merr bark against breast cancer cell lines
14.50-15.00	39/F3	La Ode Sumarlin	STUDY ACTIVITIES INHIBITION HEp-2 CELLS BY INDONESIAN LOCAL HONEY
15.00-15.10	41/F4	Sandra Hermanto	Isolation and Purification of Angiotensin Converting Enzyme Inhibitory Peptides Derived from Soy milk Hydrolysates
15.10-15.20	72/F5	Suryani	Isolation And Molecular Identification Of Lactic Acid Bacteria (Lab) In Coconut Milk Fermentation
16.00-16.10	78/F6	Imelia dewi	Utilization of Ecoenzyme Citrus reticulata in microbial fuel cell as a new potential of renewable energy
16.10-16.20	92/F7	Hira Helwati	Active edible film from Dioscorea hispida Dennst starch-chitosan composite containing ascorbic acid and turmeric extract
16.20-	114/F8	Nikmatia	Evaluation of Dissolution Profiles of Bromelain from

IPB University
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International Conference of the Indonesian Chemical Society ICICS 2019

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Presenter

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THE 8th INTERNATIONAL CONFERENCE OF THE INDONESIAN CHEMICAL SOCIETY (ICICS)

"Chemistry For Human Welfare"

August 6th – 7th 2019, IPB International Convention Center, Bogor - Indonesia

Himpunan Kimia Indonesia

Prof. Dr. Harno Dwi Pranowo
President of Indonesian Chemical Society

Dr. Mohamad Rafi
Chairman

Co-hosted:

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NUSA BANGSA UNIVERSITAS

ISOLATION AND MOLECULAR IDENTIFICATION OF LACTIC ACID BACTERIA (LAB) IN COCONUT MILK FERMENTATION

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ABSTRACT

Lactic Acid Bacteria (LAB) contains of bacteriocin which is peptide that has the capacity to isolate the growth of pathogenic bacteria, where in contrast is harmless for other good bacteria. LAB is found in material fermentation containing high carbohydrate and protein like coconut milk which is undergone the process becoming Virgin Coconut Oil (VCO). In the fermentation process, there were three layers formed; oil, blondo, and water (waste). The LAB isolation on coconut milk fermentation used MRSA + 0,5% $CaCO_3$ as the selective media. with the dilution from 10^{-1} to 10^{-7}. Here, each sample was taken from each layer formed in the milk fermentation process. The identification was carried out in two ways, first was morphology identification, and the second one was molecular identification applying the PCR method. There were 97 isolates obtained from oil layer, 23 isolates from Blondo layer, and 14 isolates from water layer. After being identified well based on both morphology, and molecular on the oil layer, there were six LAB found, which were *Lactobacillus paracasei, Lactobacillus plantarum, Micrococcus luteus, Corineaebacterium bovis, Lactobacillus thermobacterium* dan *Corineaebacterium xerocis.* Three types of LAB within the blondo were identified as *Lactobacillus plantarum, Lactobacillus paracasei* and *Lactobacillus thermobacterium.*

Keywords: Isolation, molecular identification, Virgin Coconut Oil (VC)_, Lactic Acid Bacteria (LAB), PCR

ABSTRAK

Bakteri Asam Laktat (BAL) mengandung bakteriosin yaitu peptida yang mempunyai kemampuan menghambat pertumbuhan bakteri patogen, tetapi tidak berbahaya bagi bakteri baik. Bakteri asam laktat terdapat pada fermentasi bahan yang mengandung karbohidrat dan protein tinggi seperti santan diproses menjadi Virgin Coconut Oil. Pada proses fermentasi santan menjadi Virgin Coconut Oil (VCO) terbentuk tiga lapisan yaitu lapisan Minyak, lapisan Blondo dan lapisan Air (kotoran). Isolasi Bakteri Asam Laktat dari fermentasi santan menggunakan media selektif MRSA + 0,5% $CaCO_3$ dan media MRSA saja dengan pengenceran 10^{-1} sampai 10^{-7}. Dimana sampel diambil dari setiap lapisan hasil proses fermentasi santan menjadi VCO. Identifikasi dilakukan dengan dua cara yaitu identifikasi morfologi dan identifikasi molekular dengan menggunakan metoda PCR. Dari lapisan Minyak didapat 97 isolat, lapisan blondo 23 isolat dan lapisan Air 14 isolat. Setelah diidentifikasi baik secara morfologi maupun molekular ternyata pada lapisan minyak terdapat 6 jenis Bakteri Asam Laktat (BAL) yaitu *Lactobacillus paracasei, Lactobacillus plantarum, Micrococcus luteus, Corineaebacterium bovis, Lactobacillus thermobacterium* dan *Corineaebacterium xerocis.* Pada blondo didapatkan 3 jenis Bakteri Asam Laktat (BAL) yaitu *Lactobacillus plantarum, Lactobacillus paracasei* dan *Lactobacillus thermobacterium.*

Kata kunci: isolasi, identifikasi molekular, Virgin Coconut Oil(VCO), Bakteri Asam Laktat (BAL), PCR

INTRODUCTION

Lactid acid bacteria (LAB) are bacteria isolated from materials which are rich mainly in carbohydrates and containing high protein [1] and are able to ferment those material in order to produce lactid acid. These bacteria are beneficial as source of probiotic [2],[3],[4] and contain of bacteriocin [5],[6] which is peptides that can destroy the wall of patogen bacteria cell and kill those bacteria, which is in contrast to good bacteria.

Bacteriocin has huge potential as food preservative [7], [8] besides its ablility as antimicrobial [9], [10] which in this work proves that the bacteriocin existing in lactid acid *Lactobacillus* spp can inhibit the growth of *chloramfenikol*, *Ampisilin* and *Tetrasiklin* antibiotics. In general, LAB isolation is in line with the observation of its antibacteria [10], [11], [12].

The ability of LAB containing bacteriocin which function as either antimicrobial or antibiotics is equally important with the ability of natural antibiotics which are isolated from the plants like *Trichomanes chinense* [13], and is able to inhibit *Staphilococcus aureus, Salmonella typhimurium,* and *Escherichia coli*. Other antibiotics derived from Marine Actinomycetes microbe which is *Sterptomyces sp* A11 has been determined as well [14]. Apparently, this compound can also inhibit *Bacillus substillis, E.coli,* and *Pseudomonas aeruginosa*.

Several of LAB have been isolated from various sources like in Turkey [11] obtaining 45 LAB isolates which are isolated from “Boza” sample, and consisted of *Lactococcus lactis*

subsp, *Leuconostoc citreum, Lactobacillus brevi, Lactobacillus plantarum, Lactobacillus paraplantarum, Enterococcus faeciu, Lactobacillus graminis, Pediococcus species and Lactobacillus paracasei* subsp*. paracasei*. The LAB of Medicinal herbs originated from Pirandai have also been isolated [15], and *Lactobacillus acidophillus* and *Lactococcus rafinolactis* were obtained. Traditional fermented food originated from West Sichuan Area like yoghurt is also containing LAB [16] namely *Lactobacillus* and *Lactococcus,* which is further identified applying PCR, and turned out as *Lactobacillus fermentum* and *Lactococcus lactis.* Fermented food from other areas like "Teff" contains LAB [12] such as *Lactobacillus brevis, Lactobacillus paracasei* and *Enterococcus faccium.* Besides that Dairy fermented food is also included into those which contain lactid acid bacteria *Lactobacillus* spp [10]. According to what have been mentioned above, the lactid acid bacteria isolation have been performed from various material [2] i.e., cheese, kefir grains, milk, beverage, source of poultry, cow rumen fluids, human feces, chicken feed, beef dadih, pineapple waste, and etc.

Nevertheless, there is none performed in isolating the lactid acid bacteria of the coconut milk fermentation process forming Virgin Coconut Oil (VCO). The fermented coconut milk producing VCO has many advantages such as losing weight, reducing cholesterol level, and inhibiting the groth of pathogenic bacteria, or fucntioning as both antibacteria and antivirus [17], [18], [19].

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EXPERIMENTAL SECTION

This research was conducted at several laboraties, i.e., LLDIKTI Region X Laboratory, Baso Veterinary Laboratory, Biomedical Laboratory, Medical Faculty of Andalas University. and Chemistry Laboratory of Muhammadiyah University of West Sumatra.

Materials

The materials used in this research were as follows: Coconut milk which was processed into Virgin Coconut Oil, oil layer (VCO), Blondo layer and water layer obtained from the process of making VCO through coconut milk fermentation. Whereas the media taken for isolating Lactic Acid Bacteria were Mannosa Rogosa Sharpe Broth / MRSB (Merck), CaCO3 (technical), Agarosa (Merck). In addition, sterile saline solution, MRSA and MRSA + 0.5% CaCO3 were used as the ingredients to identify morphologically, and to perform biochemical tests such as Complex Iodine, Safranine, Alcohol, Aquades and so on. Whilst the materials used for molecular identification were primary, 500µl Tris EDTA (TE), ammonium acetate, SDS-Polyacrylamide (SDS-PAGE) gel material 18-20%, comassife blue for. Ase RNA 3 µl, 70% ethanol, lysozyme, obtained ddH2O 27 µl, phenol, SDS (Sodium Dodesil Sulfate), chloroform, Proteinase K (10 mg / µl), isoamil alcohol 25: 24: 1, and protein marker with the

size of 10,000-40,000. Typically, DNA α, Tris HCl pH 8, isopropanol to fractionate DNA, ethidium bromide, agaros gel, 3M acetate, agarose, TBE buffer (Tris-Boric-EDTA) were taken.

Instrumentation

Here in this research, the equipment used in order to isolate and to conduct morphology identification as well as to perform biochemical tests of Lactic Acid Bacteria besides those commonly used glass tools were Autocklaf, Laminar Flow and Microscopes. In another hand, for molecular identification Electrophoresis was also used other than PCR.

Procedure

Isolation of Lactic Acid Bacteria

In order to get isolates, BAL was isolated from three existing layers containing coconut milk fermentation process to become Virgin Coconut Oil, namely oil layer, blondo layer and water layer. Isolation was carried out using 2 media, they were MRSA media and MRSA + media 0.5% $CaCO_3$. By applying a dilution method up to 10-7, the isolation process was performed for several times using the pour plate and streak plate method, so that a number of isolates could be obtained which was followed by the identification process morphologically along with biochemical tests.

Morphological identification

Then the obtained isolates were proceeded by performing identification morphologically where isolates were planted in MRSA media, and incubated at 37 0 C. Observed on the shape of the colony, some were convex, whereas some others were flat or concave. Examined also the color of the colonies, where there were white in color, yellow, yellowish or clear and so on. The arrangement of cells were also significant to be taken into account whether the shape of the cell was round or hollow.

Biochemical tests

The conducted biochemical tests were Catalase test, carbohydrate fermentation, oxidase, ammonia production (NH_3), and TSIA test, according to Mac Faddin (1983) procedure, then compared them to the manual (Cowan, 1975)

Molecular identification.

Molecular identification was initiated with the genomic isolation stage of lactic acid bacterial DNA isolates obtained from morphological identification and biochemical tests, then 16S rRNA gene amplification PCR 16S rDNA amplification per reaction of 30 µL using 27F

primer (5'-AGAGTTTGATCCTGGCTCAG-3 ') position 8-27 on the E. Coli chromosome and primer 1492R (5'-GGTTACCTTGTTACGA CTT -3 ') in positions of 1510-1492 on the E. coli chromosome (Nikolova et al. 2009), followed by analyzing it in the Electrophoresis Gel and was ended by performing its sequencing data analysis.

RESULT AND DISCUSSION

Isolation of Lactic Acid Bacteria

The colony of Lactic Acid Bacteria isolation using the MRSA media in the absence of adding 0,5% $CaCO_3$ was able to be measured at the dilution of 10^{-5} up to 10^{-7}. At 10^{-1} up to 10^{-4} dilution, the colonies were grown so dense therefore they were unmeasureable.The grown bacteria were yellowish, convex, and rather shiny. How ever, as highlighted by Delfaedah and Sumaryati Syukur (2013); Hoque (2010), Heravi (2011), Syukur and Husmaini (2014) that those grown colonies were identical among each other, and there were co certainty that these were LAB colonies. To confirm that these were LAB colonies, Husmaini and Endang Purwati (2012) suggested to proceed using KIT AOI CHL 50. The LAB colonies which were grown in MRSA media can be seen in Figure 1 below:

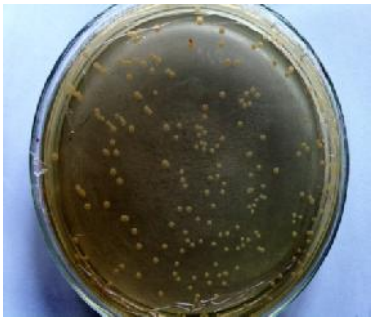


Figure 1. LAB isolates colony grown in MRS media

LAB isolation process using MRSA media + 0,5% $CaCO_3$, would form the "Halo" area, where one colony was found in its center, and was confirmed as LAB bacteria colony. $CaCO_3$ would be reacted with the acid produced by LAB, neutralized it, and made the area free from bacteria and in clear condition. The isolates grown in the center of "Halo" area were picked using ose, and then scratch them to MRSA media for morphology identification. LAB colony grown in MRSA media + 0,5% $CaCO_3$ can be seen in Figure 2. below:

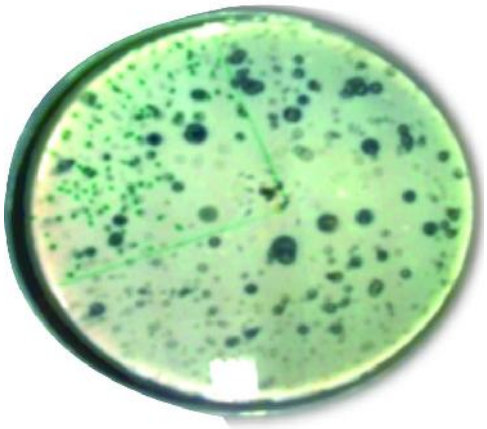


Figure 2. LAB isolate colony grown in MRSA media + 0,5% $CaCO_3$

The use of MRSA media + 0,5% $CaCO_3$ in isolating LAB was in accordance as highlighted by Rukmini Putri (2012); where she fermented Lactid Acid Bacteria obtained from fermentation of *Growol*, Indonesia traditional cuisine, by using MRSA media added with 1,5 % $CaCO_3$. Whereas, Nguyen (2010_, isolated LAB using MRSA media 1 % $CaCO_3$ as well on the Vietnamese traditional food namely *Nem chua*, and Sarkono (2010) isolated the LAB derived from abalone using MRSA media + 0,5 % $CaCO_3$, the indicated colony was the growth of LAB bacteria marked by the clear zone in its surrounding.

The LAB isolate colony after being measured is shown in Table 1 below:

Table 1. LAB isolates amount of isolated result

No.	Layer	Isolate Amount
1.	Oil	97
2.	Blondo	23
3.	Water	14

Morphology Identification

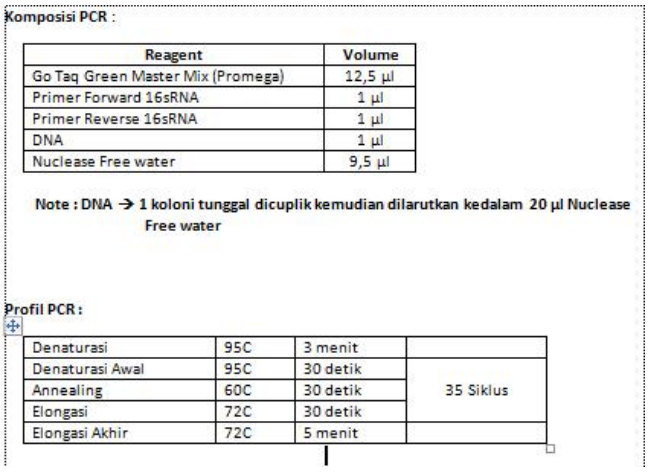
There were 134 LAB isolates of the isolation result using the MRSA media + $CaCO_3$ would experience further analysis process on morphology identification. It resulted variation in shape of colony like convex, flat, and concave, variation in color like white, yellow, yellowish, and clear, and variation in smell like odour and odourless. This morphology identification result of LAB isolates was confirmed by biochemical test's results such as gram staining, motility test, and others, and combined with physiology test as shown in Table 2 below:

Table 2. Result of Morphology Identification on LAB Isolates

No.	Layer	Type of LAB
1.	**Oil Layer**	*Lactobacillus plantarum* *Lactobacillus paracasei* *Micrococcus luteus* *Corineaebacterium bovis* *Corineaebacterium xerosis* *Lactobacillus thermobacterium*
2.	**Blondo Layer**	*Lactobacillus plantarum* *Lactobacillus paracasei* *Lactobacillus thermobacterium*
3.	**Water Layer**	*Micrococcus luteus* *Corineaebacterium bovis* *Corineaebacterium xerosis*

Molecular Identification

From 134 LAB isolates obtained, some were performed further identification molecularly using PCR, in order to determine their LAB types, and to find out the sequence of its LAB DNA. The molecular identification was initially conducted onto four isolates. Its analysis result using the PCR composition and profil was pointed out in the Figure 3 below:



Komposisi PCR :

Reagent	Volume
Go Taq Green Master Mix (Promega)	12,5 µl
Primer Forward 16sRNA	1 µl
Primer Reverse 16sRNA	1 µl
DNA	1 µl
Nuclease Free water	9,5 µl

Note : DNA → 1 koloni tunggal dicuplik kemudian dilarutkan kedalam 20 µl Nuclease Free water

Profil PCR :

Denaturasi	95C	3 menit	
Denaturasi Awal	95C	30 detik	35 Siklus
Annealing	60C	30 detik	
Elongasi	72C	30 detik	
Elongasi Akhir	72C	5 menit	

Figure 3. PCR Composition and Profile

Produced electropherogram as shown in Figure 4 below:

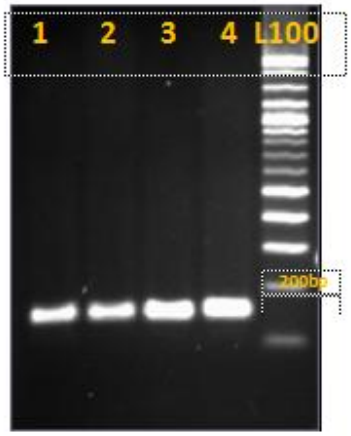


Figure 4. Electropherogram on PCR Result using **16s RNA Primer**

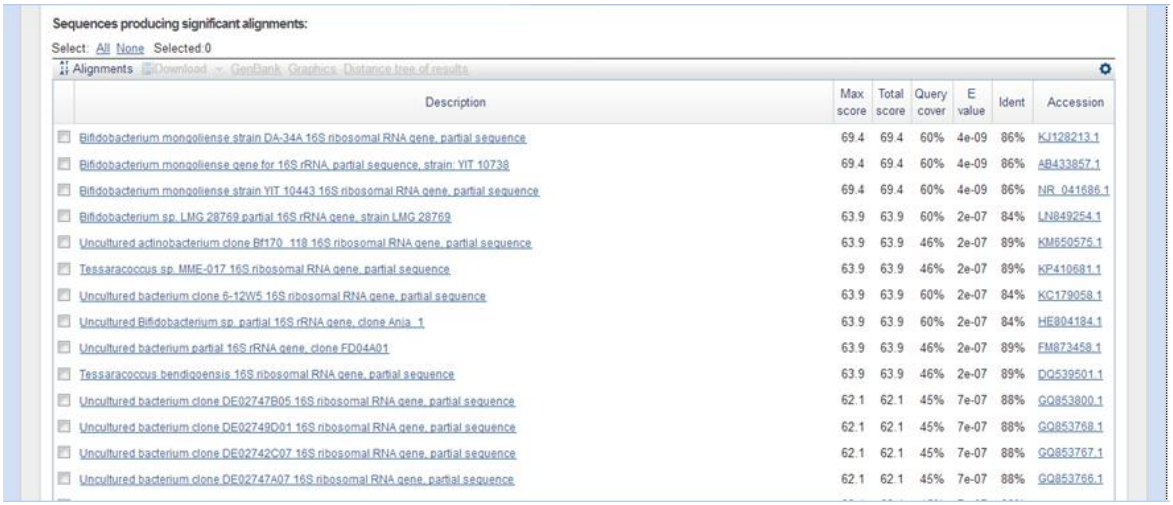
Its sequencing is highlited in the Figure 5 below:

1. **Bac1**

   Forward :

   GACGTCCCATGAGAGTTTGTACAGCCGAAGCCGGTGGCCTAACCTTTTTGGGGAGAGCCCC

   CTAAAGCGTGAGACATGAGAGGGGGGAGATCTCATAAAGGTGTCCGTAAAA



Sequences producing significant alignments:

Select: All None Selected:0

Alignments Download GenBank Graphics Distance tree of results

Description	Max score	Total score	Query cover	E value	Ident	Accession
Bifidobacterium mongoliense strain DA-34A 16S ribosomal RNA gene, partial sequence	69.4	69.4	60%	4e-09	86%	KJ128213.1
Bifidobacterium mongoliense gene for 16S rRNA, partial sequence, strain: YIT 10738	69.4	69.4	60%	4e-09	86%	AB433857.1
Bifidobacterium mongoliense strain YIT 10443 16S ribosomal RNA gene, partial sequence	69.4	69.4	60%	4e-09	86%	NR_041686.1
Bifidobacterium sp. LMG 28769 partial 16S rRNA gene, strain LMG 28769	63.9	63.9	60%	2e-07	84%	LN849254.1
Uncultured actinobacterium clone Bf170_118 16S ribosomal RNA gene, partial sequence	63.9	63.9	46%	2e-07	89%	KM650575.1
Tessaracoccus sp. MME-017 16S ribosomal RNA gene, partial sequence	63.9	63.9	46%	2e-07	89%	KP410681.1
Uncultured bacterium clone 6-12W5 16S ribosomal RNA gene, partial sequence	63.9	63.9	60%	2e-07	84%	KC179058.1
Uncultured Bifidobacterium sp. partial 16S rRNA gene, clone Ania_1	63.9	63.9	60%	2e-07	84%	HE804184.1
Uncultured bacterium partial 16S rRNA gene, clone FD04A01	63.9	63.9	46%	2e-07	89%	FM873458.1
Tessaracoccus bendigoensis 16S ribosomal RNA gene, partial sequence	63.9	63.9	46%	2e-07	89%	DQ539501.1
Uncultured bacterium clone DE02747B05 16S ribosomal RNA gene, partial sequence	62.1	62.1	45%	7e-07	88%	GQ853800.1
Uncultured bacterium clone DE02749D01 16S ribosomal RNA gene, partial sequence	62.1	62.1	45%	7e-07	88%	GQ853768.1
Uncultured bacterium clone DE02742C07 16S ribosomal RNA gene, partial sequence	62.1	62.1	45%	7e-07	88%	GQ853767.1
Uncultured bacterium clone DE02747A07 16S ribosomal RNA gene, partial sequence	62.1	62.1	45%	7e-07	88%	GQ853766.1

Figure 5. Result of Isolate No 1 Sequencing

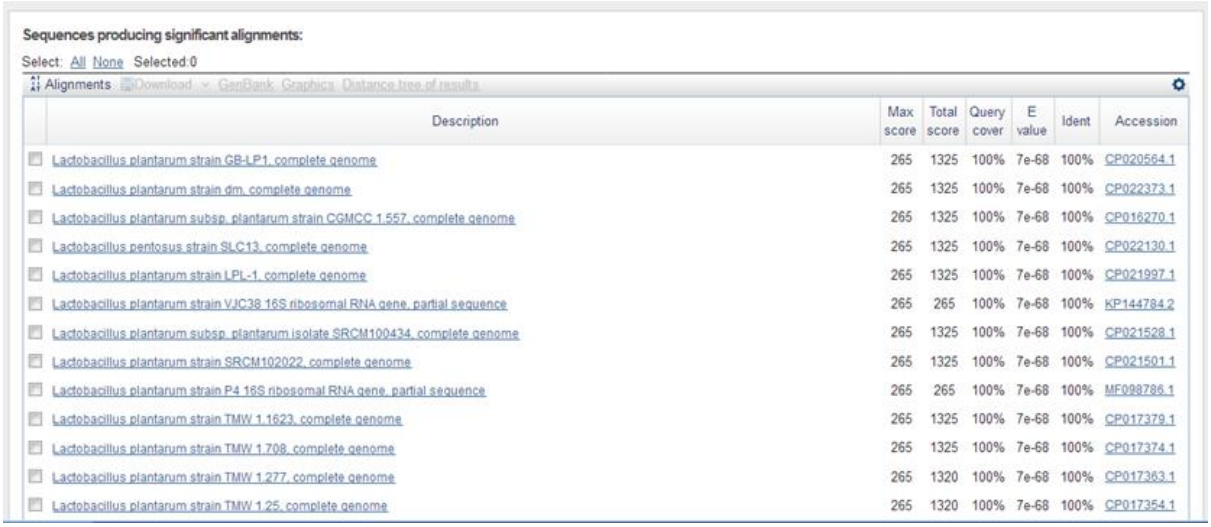
In the figure shown above, the sequencing result on LAB No 1 isolate is not detected.

The following is Figure 6. The isolate No 2 sequencing result.

2. **Bac2**

Consensus :

CGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACACCATGAGAGTTTGTAACACC
CAAAGTCGGTGGGGTAACCTTTTAGGAACCAGCCGCCTAAGGTGGGACAGATGATTAGGG
TGAAGTCGTAACAAGGTAGCC



Sequences producing significant alignments:

Select: All None Selected:0

Alignments Download GenBank Graphics Distance tree of results

Description	Max score	Total score	Query cover	E value	Ident	Accession
Lactobacillus plantarum strain GB-LP1, complete genome	265	1325	100%	7e-68	100%	CP020564.1
Lactobacillus plantarum strain dm, complete genome	265	1325	100%	7e-68	100%	CP022373.1
Lactobacillus plantarum subsp. plantarum strain CGMCC 1.557, complete genome	265	1325	100%	7e-68	100%	CP016270.1
Lactobacillus pentosus strain SLC13, complete genome	265	1325	100%	7e-68	100%	CP022130.1
Lactobacillus plantarum strain LPL-1, complete genome	265	1325	100%	7e-68	100%	CP021997.1
Lactobacillus plantarum strain VJC38 16S ribosomal RNA gene, partial sequence	265	265	100%	7e-68	100%	KP144784.2
Lactobacillus plantarum subsp. plantarum isolate SRCM100434, complete genome	265	1325	100%	7e-68	100%	CP021528.1
Lactobacillus plantarum strain SRCM102022, complete genome	265	1325	100%	7e-68	100%	CP021501.1
Lactobacillus plantarum strain P4 16S ribosomal RNA gene, partial sequence	265	265	100%	7e-68	100%	MF098786.1
Lactobacillus plantarum strain TMW 1.1623, complete genome	265	1325	100%	7e-68	100%	CP017379.1
Lactobacillus plantarum strain TMW 1.708, complete genome	265	1325	100%	7e-68	100%	CP017374.1
Lactobacillus plantarum strain TMW 1.277, complete genome	265	1320	100%	7e-68	100%	CP017363.1
Lactobacillus plantarum strain TMW 1.25, complete genome	265	1320	100%	7e-68	100%	CP017354.1

Sequencing result on Isolate No 2

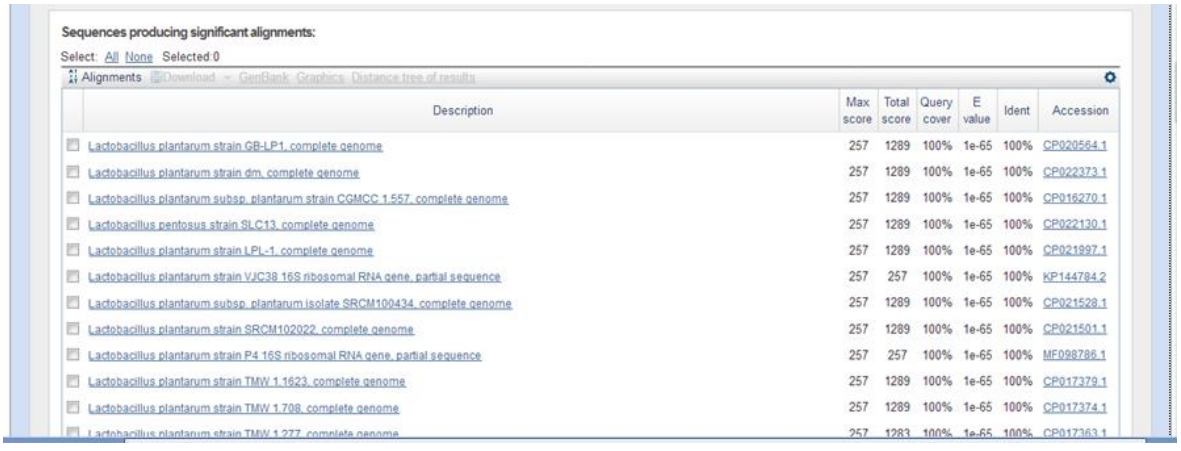
Presented in Figure 6 that the isolate is *Lactobacillus plantarum*

The following is Figure 7. containing the sequencing result of LAB No 3,

3. **Bac3**

Consensus :

GAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACACCATGAGAGTTTGTAACACCCAAA
GTCGGTGGGGTAACCTTTTAGGAACCAGCCGCCTAAGGTGGGACAGATGATTAGGGTGAA
GTCGTAACAAGGTAGCC



Sequences producing significant alignments:

Select: All None Selected:0

Alignments Download GenBank Graphics Distance tree of results

Description	Max score	Total score	Query cover	E value	Ident	Accession
Lactobacillus plantarum strain GB-LP1, complete genome	257	1289	100%	1e-65	100%	CP020564.1
Lactobacillus plantarum strain dm, complete genome	257	1289	100%	1e-65	100%	CP022373.1
Lactobacillus plantarum subsp. plantarum strain CGMCC 1.557, complete genome	257	1289	100%	1e-65	100%	CP016270.1
Lactobacillus pentosus strain SLC13, complete genome	257	1289	100%	1e-65	100%	CP022130.1
Lactobacillus plantarum strain LPL-1, complete genome	257	1289	100%	1e-65	100%	CP021997.1
Lactobacillus plantarum strain VJC38 16S ribosomal RNA gene, partial sequence	257	257	100%	1e-65	100%	KP144784.2
Lactobacillus plantarum subsp. plantarum isolate SRCM100434, complete genome	257	1289	100%	1e-65	100%	CP021528.1
Lactobacillus plantarum strain SRCM102022, complete genome	257	1289	100%	1e-65	100%	CP021501.1
Lactobacillus plantarum strain P4 16S ribosomal RNA gene, partial sequence	257	257	100%	1e-65	100%	MF098786.1
Lactobacillus plantarum strain TMW 1.1623, complete genome	257	1289	100%	1e-65	100%	CP017379.1
Lactobacillus plantarum strain TMW 1.708, complete genome	257	1289	100%	1e-65	100%	CP017374.1
Lactobacillus plantarum strain TMW 1.277, complete genome	257	1283	100%	1e-65	100%	CP017363.1

Figure 7. The isolate sequencing result on LAB No 3

Presented in Figure 7, it can be seen that LAB isolate of sample No 3 is *Lactobacillus plantarum*

The sequencing result of isolate sample No 4 is pointed out in Figure 8 below,

4. **Bac4**

Consensus:

CGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACACCATGAGAGTTTGTAACACC
CGAAGCCGGTGGCGTAACCCTTTTAGGGAGCGAGCCGTCTAAGGTGGGACAAATGATTAG
GGTGAAGTCGTAACAAGGTAGCCGTAA

Sequences producing significant alignments:

Select: All None Selected:0

Alignments Download GenBank Graphics Distance tree of results

Description	Max score	Total score	Query cover	E value	Ident	Accession
Bacterium strain A2 16S ribosomal RNA gene, partial sequence	276	276	100%	3e-71	100%	KX268350.1
Lactobacillus paracasei strain HD1.7 16S ribosomal RNA gene, partial sequence	276	276	100%	3e-71	100%	EU249147.1
Lactobacillus rhamnosus strain L156.4 16S ribosomal RNA gene, partial sequence	274	274	99%	1e-70	100%	KX644947.1
Lactobacillus casei strain LC5, complete genome	274	1372	99%	1e-70	100%	CP017065.1
Lactobacillus rhamnosus strain 4B15, complete genome	274	1372	99%	1e-70	100%	CP021426.1
Lactobacillus paracasei strain IIA, complete genome	274	1372	99%	1e-70	100%	CP014985.1
Lactobacillus rhamnosus strain Pen, complete genome	274	1372	99%	1e-70	100%	CP020464.1
Lactobacillus rhamnosus strain WQ2 genome	274	274	99%	1e-70	100%	CP020016.1
Lactobacillus rhamnosus strain BFE5264, complete genome	274	1372	99%	1e-70	100%	CP014201.1
Lactobacillus casei strain K1C 16S ribosomal RNA gene, partial sequence; 16S-23S ribosomal RNA intergenic spacer, complete sequence; and 23S	274	274	99%	1e-70	100%	KU954559.1
Lactobacillus rhamnosus strain LRB, complete genome	274	1372	99%	1e-70	100%	CP016823.1
Lactobacillus rhamnosus gene for 16S ribosomal RNA, partial sequence, strain: NGRI 0110	274	274	99%	1e-70	100%	LC177236.1
Lactobacillus paracasei strain FS3 16S ribosomal RNA gene, partial sequence	274	274	99%	1e-70	100%	KU315084.1
Lactobacillus rhamnosus strain ASCC 290 genome	274	274	99%	1e-70	100%	CP014645.1

Figure 8. The isolate sequencing result on LAB No 4

It is identified within Figure 8 that the isolate of sample No 4 is *Lactobacillus paracasei*

Then, molecular identification was performed with the second four isolates by conducting the analysis of these four isolates consecutively. For the bacteria DNA of PCR amplification result on 165 ribosomal sequencing zone is highlighted in the following Figure 9.,

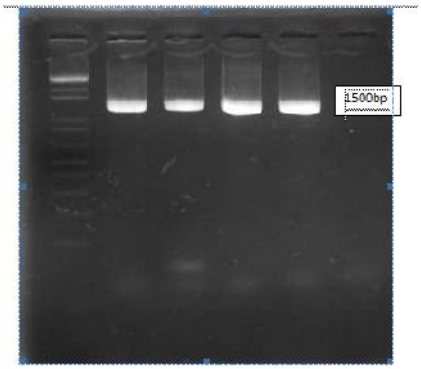


Figure 9. Electrophoresis result on 4 isolates; M0. A5 , M16.16.2 and M16.4

Picture Description:

- The sample of PCR product is in the size of 1500bp, (-) control negative
- Ladder DNA 1kb plus 100, 200, 300, 400, 500, 650, 850, 1000, 1650, 2000, 3000, 4000 bp.
    - Fasta format was obtained from the result of sample sequencing analysis using Bioedit program. The fasta format of the sample is as follow:

>CONTIQ_M16.4_1440bp_*Lactobacillus plantarum*_100%

GCTCAGGACGAACGCTGGCGGCGTGCCTAATACATGCAAGTCGAACGAACTCTGGTATTGATTGGTGCTT
GCATCATGATTTACATTTGAGTGAGTGGCGAACTGGTGAGTAACACGTGGGAAACCTGCCCAGAAGCGGGGGATA
ACACCTGGAAACAGATGCTAATACCGCATAACAACTTGGACCGCATGGTCCGAGCTTGAAAGATGGCTTCGGCTAT
CACTTTTGGATGGTCCCGCGGCGTATTAGCTAGATGGTGGGGTAACGGCTCACCATGGCAATGATACGTAGCCGAC
CTGAGAGGGTAATCGGCCACATTGGGACTGAGACACGGCCCAAACTCCTACGGGAGGCAGCAGTAGGGAATCTTC
CACAATGGACGAAAGTCTGATGGAGCAACGCCGCGTGAGTGAAGAAGGGTTTCGGCTCGTAAAACTCTGTTGTTA
AAGAAGAACATATCTGAGAGTAACTGTTCAGGTATTGACGGTATTTAACCAGAAAGCCACGGCTAACTACGTGCCA
GCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGATTTATTGGGCGTAAAGCGAGCGCAGGCGGTTTTTA
AGTCTGATGTGAAAGCCTTCGGCTCAACCGAAGAAGTGCATCGGAAACTGGGAAACTTGAGTGCAGAAGAGGACA
GTGGAACTCCATGTGTAGCGGTGAAATGCGTAGATATATGGAAGAACACCAGTGGCGAAGGCGGCTGTCTGGTCT
GTAACTGACGCTGAGGCTCGAAAGTATGGGTAGCAAACAGGATTAGATACCCTGGTAGTCCATACCGTAAACGAT
GAATGCTAAGTGTTGGAGGGTTTCCGCCCTTCAGTGCTGCAGCTAACGCATTAAGCATTCCGCCTGGGGAGTACGG
CCGCAAGGCTGAAACTCAAAGGAATTGACGGGGGCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCTAC
GCGAAGAACCTTACCAGGTCTTGACATACTATGCAAATCTAAGAGATTAGACGTTCCCTTCGGGGACATGGATACA
GGTGGTGCATGGTTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCTTATTATC
AGTTGCCAGCATTAAGTTGGGCACTCTGGTGAGACTGCCGGTGACAAACCGGAGGAAGGTGGGGATGACGTCAAA
TCATCATGCCCCTTATGACCTGGGCTACACACGTGCTACAATGGATGGTACAACGAGTTGCGAACTCGCGAGAGTA
AGCTAATCTCTTAAAGCCATTCTCAGTTCGGATTGTAGGCTGCAACTCGCCTACATGAAGTCGGAATCGCTAGTAAT
CGCGGATCAGCATGCCGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACACCATGAGAGTTTGTAAC
ACCCAAAGTC

>CONTIQ_A5_1430bp_*Lactobacillus plantarum*_100%

ACGCTGGCGGCGTGCCTAATACATGCAAGTCGAACGAACTCTGGTATTGATTGGTGCTTGCATCATGATTT
ACATTTGAGTGAGTGGCGAACTGGTGAGTAACACGTGGGAAACCTGCCCAGAAGCGGGGGATAACACCTGGAAA
CAGATGCTAATACCGCATAACAACTTGGACCGCATGGTCCGAGCTTGAAAGATGGCTTCGGCTATCACTTTTGGATG
GTCCCGCGGCGTATTAGCTAGATGGTGGGGTAACGGCTCACCATGGCAATGATACGTAGCCGACCTGAGAGGGTA
ATCGGCCACATTGGGACTGAGACACGGCCCAAACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCACAATGGACG
AAAGTCTGATGGAGCAACGCCGCGTGAGTGAAGAAGGGTTTCGGCTCGTAAAACTCTGTTGTTAAAGAAGAACAT
ATCTGAGAGTAACTGTTCAGGTATTGACGGTATTTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGCCGCGGT
AATACGTAGGTGGCAAGCGTTGTCCGGATTTATTGGGCGTAAAGCGAGCGCAGGCGGTTTTTTAAGTCTGATGTGA
AAGCCTTCGGCTCAACCGAAGAAGTGCATCGGAAACTGGGAAACTTGAGTGCAGAAGAGGACAGTGGAACTCCAT
GTGTAGCGGTGAAATGCGTAGATATATGGAAGAACACCAGTGGCGAAGGCGGCTGTCTGGTCTGTAACTGACGCT
GAGGCTCGAAAGTATGGGTAGCAAACAGGATTAGATACCCTGGTAGTCCATACCGTAAACGATGAATGCTAAGTG
TTGGAGGGTTTCCGCCCTTCAGTGCTGCAGCTAACGCATTAAGCATTCCGCCTGGGGAGTACGGCCGCAAGGCTGA
AACTCAAAGGAATTGACGGGGGCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCTACGCGAAGAACCTT
ACCAGGTCTTGACATACTATGCAAATCTAAGAGATTAGACGTTCCCTTCGGGGACATGGATACAGGTGGTGCATGG
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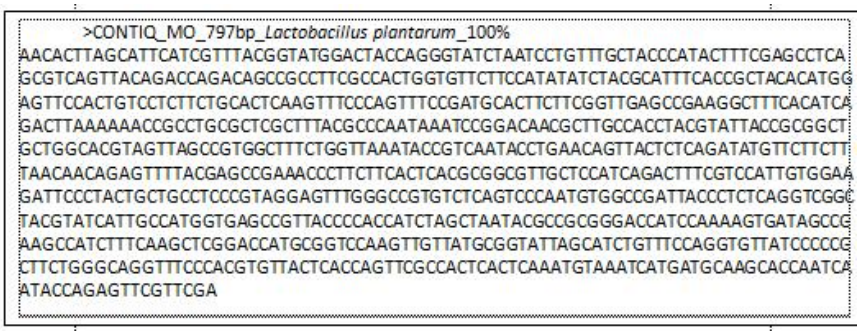

The sequence molecular identification result of the second sample on DNA 4 is shown in Table 2. as follow:

Table 2. Result of LAB molecular identification

No.	Isolate Code	LAB Type
1.	**IsolatMO**	*Lactobacillus plantarum*
2.	**IsolatM16.16.2**,	*Lactobacillus plantarum*
3.	**IsolatA5**	*Lactobacillus plantarum*
4.	**Isolat16.4**,	*Lactobacillus plantarum*

Phylogenetics Analysis

In the following four samples, phylogenetics analysis was carried out using the **bootsrap method** as seen in Figure 10 below:

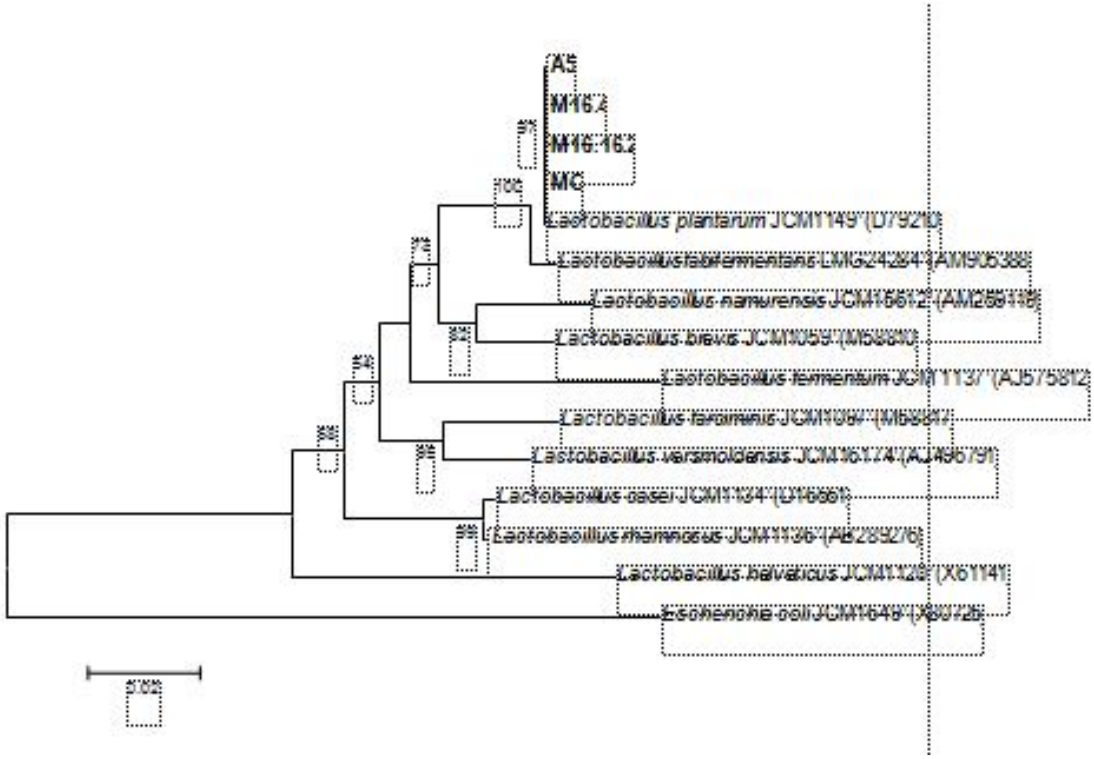

Figure 10. Phylogenetic tree obtained from Neighbor Joining analysis with 1000 repetition

CONCLUSION

Regarding to the research conducted and the results obtained, it can be concluded that:

1. A total of 134 isolates of BAL (Lactic Acid Bacteria) can be isolated from the fermentation process of coconut milk consisting of 97 isolates deriving from the oil layer, 23 isolates deriving from Blondo and 14 isolates deriving from the water layer.
2. There are six Lactic Acid bacteria that can be identified morphologically, namely *Lactobacillus plantarum, Lactobacillus paracasei, Lactobacillus thermobacterium, Micrococcus luteus, Corineaebacterium bovis dan Corineaebacterium xerocis.*
3. The molecularly identified Lactic Acid Bacteria using PCR is broken down into three types, namely Lactobacillus plantarum, *Lactobacillus plantarum* strain JCM1149T, and *Lactobacillus paracasei.*

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This research consumed huge amount of work, and dedication. It would not have been possible without having support of many individuals and organizations. Therefore we would like to extend our sincere gratitude to all of them.

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3. We would like to express our sincere thanks towards the Head of Biochemistry at Andalas University.
4. Nevertheless, we express our gratitude toward Head of Laboratory of Region X Private Higher Education Coordination (Kopertis).